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# SENSITIVITY OF ACTION POTENTIAL TO CHANGES OF INWARD RECTIFIER POTASSIUM CURRENT IK1 IS DIFFERENT IN RECENT MODELS OF HUMAN VENTRICULAR CARDIOMYOCYTES

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**Abstract:** The inwardly rectifying potassium current  $I_{K1}$  is one of the principle ionic currents responsible for repolarization phase of mammalian action potentials (APs). To estimate the impact of individual ionic currents on AP configuration, mathematical models have been widely used. In this study, we compare the effects of alcohol-induced changes of  $I_{K1}$  on AP duration (APD) as simulated in four recently published computer models of human ventricular cells. As expected, increasing or decreasing  $I_{K1}$  conductance by 20% respectively caused a shortening or a lengthening of APD. However, the effect was largely model-dependent, ranging from 1% to about 15% change of APD. Given the conflicting available experimental data on the features of  $I_{K1}$  in human ventricular myocytes there is a need for a set of well-established end-point constraints for a reliable human ventricular myocyte model to be generated.

Keywords: Cardiac cell, Action potential, Inward rectifier potassium current, Quantitative modelling.

## 1. Introduction

In our recent work, we studied the effects of ethanol on the inward rectifier potassium current  $(I_{K1})$  in adult rat ventricular myocytes (Bébarová et al., 2013). The results showed that ethanol affects  $I_{K1}$  in dual ways; it causes an inhibition of  $I_{K1}$  at very low concentrations up to 0.8 mM (equivalent to ~0.37‰ of ethanol in the blood) and an increase at concentrations above 20 mM (equivalent to ~0.92‰ of ethanol in the blood). To simulate the functional consequences of these changes of  $I_{K1}$  on human cardiac cells we decided to use our own and three other recently published models of human ventricular myocytes (Hrabcová et al., 2013; O'Hara et al., 2011; Fink et al., 2008; Iyer et al., 2004) for comparison. Surprisingly, changes of  $I_{K1}$  led to substantially different effects on AP in these four models indicating different sensitivities of the models to  $I_{K1}$  variations.

## 2. Methods

To compare the sensitivity of action potential (AP) to changes of  $I_{K1}$  in recently published models selected for this study (Hrabcová et al., 2013; O'Hara et al., 2011; Fink et al., 2008; Iyer et al., 2004), we performed simulations of APs and  $I_{K1}$  at 1Hz stimulation at steady-state (after 300 s stimulation – control conditions) and after an increase and decrease of  $I_{K1}$ -channels conductivity ( $g_{K1}$ ) by 20%. In each case, AP duration at 90% repolarisation (APD<sub>90</sub>) and relative change of APD<sub>90</sub> were evaluated.

The simulations on our model (Hrabcová et al., 2013) were performed using the computational system MATLAB 7.2 (MathWorks, Natick, MA, USA) and the solver for stiff systems ODE-15s. Simulations on the other models (O'Hara et al., 2011; Fink et al., 2008; Iyer et al., 2004) were performed using the computational environment for cellular modelling, CORv.0.9.31.1409 (Dr. Alan Garny), and the CellML codes of the models available at http://models.cellml.org/electrophysiology.

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#### 3. Results

Fig. 1 shows the simulated APs using all four models under conditions described in methods. In all cases, as expected, the 20% decrease of  $g_{K1}$  caused a reduction of  $I_{K1}$  and, consequently, a prolongation of AP duration. Analogically, the 20% increase of  $g_{K1}$  resulted in an increase of  $I_{K1}$  and subsequent shortening of AP duration. However, as evident from the figure, the potencies of the  $I_{K1}$  changes to affect AP were substantially different in these models.



Fig. 1: Simulations of APs and I<sub>K1</sub> in 1Hz steady-state (control conditions - red lines) and after decrease or increase of g<sub>K1</sub> by 20% (blue and green lines respectively) using the models of human ventricular cardiomyocytes recently published by: a) Hrabcová et al. (2013) ;b) O'Hara et al. (2011); c) Fink et al. (2008); and d) Iyer et al. (2004). APD<sub>90</sub> stands for the duration of action potential at 90% of repolarisation.

Fig. 2 shows that the highest sensitivity of AP to changes of  $I_{K1}$  was found in the model of Hrabcová et al. (2013) where the 20% decrease of  $g_{K1}$  caused a relative increase of APD<sub>90</sub> by 14.5% and the 20% increase of  $g_{K1}$  caused a relative decrease of APD<sub>90</sub> by 10.1%. On the contrary, the lowest sensitivity of AP to

changes of  $I_{K1}$  was exhibited by the model of O'Hara et al. (2011) where the same changes of  $g_{K1}$  resulted in only 1.2% increase and 1.4% decrease of APD<sub>90</sub>, respectively.



Fig. 2: Comparison of relative increase and decrease of  $APD_{90}$  (blue and green columns respectively) evoked respectively by a decrease or increase of  $g_{K1}$  by 20% from control values in the models of human ventricular cardiomyocytes recently published by Hrabcová et al. (2013), O'Hara et al. (2011), Fink et al. (2008) and Iyer et al. (2004).

### 4. Discussion and Conclusions

The prominent differences between sensitivities of APD to changes of  $I_{K1}$  in the explored models reflect the inconsistencies in mathematical description of  $I_{K1}$  properties due to the lack of experimental data from human ventricular cardiomyocytes gathered up to date. The highest sensitivity was observed in the model of Hrabcová et al. (2013) with formulation of  $I_{K1}$  based on the description in Iyer et al. (2004) while the lowest sensitivity was found in the model of O'Hara et al. (2011).

During the last decade, other comparative studies have been published in an effort to define the contribution of individual components of potassium current to repolarization of the AP. For example, Fink et al. (2006) have compared ten Tusscher et al. (2004) and Iyer et al. (2004) models and have concluded that the effects of a fixed percentage reduction of  $I_{K1}$  give rise to significantly different prolongation of AP in these two models. However, they noted that it was not possible to determine unequivocally which of these models would be more reliable for simulation of AP repolarization because reliable data on  $I_{K1}$  in human ventricle were not available in the whole range of physiological voltages. In 2008, Fink et al. reformulated  $I_{K1}$  to better reproduce the data obtained from human ventricular myocytes in their new model. Later on, Grandi et al. (2010) proposed an improved computational model of the human epicardial and endocardial myocytes, based on some of the best features of previous models combined with newer data.  $I_{K1}$  blockade increases APD rather modestly in these two later models consistently with other published experimental data (Rudy et al., 2008).

Nevertheless, the experimental results related to the properties of  $I_{K1}$  in human ventricular myocytes are not yet complete, and the sample size of available data sets is too small so far. Because of ethical reasons, it is practically impossible to study sufficient numbers of normal human cardiac cells to fully characterize their electrophysiological properties. On occasion, experimental animal models can help to fill important gaps in the missing data. They should be, however, used with caution. For example,  $I_{K1}$  changes appear to affect the APD considerably more in guinea pig (Miake et al., 2003) and dog (Jost et al., 2013) than in human cardiomyocytes.

In conclusion, the formulation of  $I_{K1}$  in the models of Fink et al. (2008) and Grandi et al. (2010) seem to be currently best adjusted to available measured experimental data from human cardiomyocytes and will likely provide the most reliable view on the effect of ethanol induced block of  $I_{K1}$  on action potential in human ventricular cells.

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## References

- Bébarová, M., Matejovič, P., Pásek, M., Šimurdová, M., Šimurda, J. (2013) Ethanol and its principle metabolite acetaldehyde affect inward rectifier potassium current IK1 in rat ventricular myocytes (abstract). Biomedical Papers of the Faculty of Medicine and Dentistry of Palacky University Olomouc, Czech Republics, 157, pp. S1.
- Fink, M., Giles, W. R., Noble, D. (2006) Contributions of inwardly rectifying K+ currents to repolarization assessed using mathematical models of human ventricular myocytes. Philosophical Transactions of the Royal Society A, 364, pp. 1207-1222.
- Fink, M., Noble, D., Virag, L., Varro, A., Giles, W. R. (2008) Contributions of HERG K+ current to repolarisation of the human ventricular action potential. Progress in Biophysics and Molecular Biology, 96, pp. 357-376.
- Grandi, E., Pasqualini, F. S., Bers, D. M. (2010) A novel computational model of the human ventricular action potential and Ca transient. Journal of Molecular and Cellular Cardiology, 48, pp. 112-121.
- Hrabcová, D., Pásek, M., Šimurda, J., Christé, G. (2013) Effect of ion concentration changes in the limited extracellular spaces on sarcolemmal ion transport and Ca2+ turnover in a model of human ventricular cardiomyocyte. International Journal of Molecular Sciences, 14, pp. 24271-24292.
- Iyer, V., Mazhari, R., Winslow, R. L. (2004) A computational model of the human left-ventricular epicardial myocyte. Biophysical Journal, 87, pp. 1507-1525.
- Jost, N., Virág, L., Comtois, P., Ordög, B., Szuts, V., Seprényi, G., Bitay, M., Kohajda, Z., Koncz, I., Nagy, N., Szél, T., Magyar, J., Kovács, M., Puskás, L. G., Lengyel, C., Wettwer, E., Ravens, U., Nánási, P. P., Papp, J. G., Varro, A., Nattel, S. (2013) Ionic mechanisms limiting cardiac repolarization reserve in humans compared to dogs. Journal of Physiology, 591, pp. 4189-4206.
- Miake, J., Marbán, E., Nuss, H. B. (2003) Functional role of inward rectifier current in heart probed by Kir2.1 overexpression and dominant-negative suppression. The Journal of Clinical Investigation, 111, pp. 1529-1536.
- O'Hara, T., Virág, L., Varró, A., Rudy, Y. (2011) Simulation of the undiseased human cardiac ventricular action potential: Model formulation and experimental validation. PLoS Computational Biology, 7, e1002061.
- Rudy, Y., Ackerman, M. J., Bers, D. M., Clancy, C. E., Houser, S. R, London, B., McCulloch, A. D., Przywara, D. A., Rasmusson, R. L., Solaro, R. J., Trayanova, N. A., Van Wagoner, D. R., Varró, A., Weiss, J. N., Lathrop, D. A. (2008) Systems approach to understanding electromechanical activity in the human heart: a national heart, lung, and blood institute workshop summary. Circulation, 118, pp. 1202-1211.
- Ten Tusscher, K. H., Noble, D., Noble, P. J., Panfilov, A. V. (2004) A model for human ventricular tissue. American Journal of Physiology, 286, pp. H1573-H1589.